

## Polyhydroxyalkanoates preparation by Fusarium moniliforme using sugarcane as substrate

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### Abstract

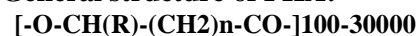
Continuous usage of conventional plastic cause waste accumulation and green house gases emission. Due to this reason development of biogreen material has been enhanced as they exert negligible side effects on environment. Biogreen materials include polylactic acid (PLA), naturally occurring zein and polyhydroxyalkanoates. Now a days, polyhydroxyalkanoates has been attracting major interest due to its compatibility with synthetic plastic. PHA is synthesized by a number of microorganisms as intracellular inclusion. *Fusarium moniliforme* is one of the microorganism that accumulate PHA using sugarcane juice as carbon substrate and ammonium ferrous sulphate as nitrogen substrate. Its production cost is comparatively less as compared to that in which glucose is used as substrate. But the overall production cost is high which is the major drawback of this method of PHA preparation. That's why efforts are going on in developing some mutant strain or using some inexpensive substrate in order to reduce the production cost.

### Introduction

Synthetic plastic has replaced glass wood and other constructional materials and even metals in much industrial, domestic and environmental application not only due to their mechanical and

thermal properties but also due to its stability and durability. But their persistence in environment causes high cost of solid waste disposal as well as the potential hazards from waste incineration such as dioxin emission from PVC resulted waste management problem. That's why recent technologies are directed towards the development of biodegradable plastic. PHAs have been attained major interest due to its similar material properties with conventional plastic and its biodegradability.

### General structure of PHA:



n=1 R = hydrogen	poly-3-hydroxypropionate
methyl	poly-3hydroxybutyrate
ethyl	poly-3-hydroxyvalerate
propyl	poly-3hydroxyhexanoate
pentyl	poly-3-hydroxyoctanoate
nonyl	poly-3-hydroxydodecanoate
n=2 R = hydrogen	poly-4-hydroxybutyrate
n=3 R = hydrogen	poly-5-hydroxyvalerate

It is known to accumulate by a number of microorganisms. *Fusarium moniliforme* a fungus is also known to accumulate PHA. It causes bakanae and foot rot disease in rice plant i.e. isolated from the infected roots of rice plant.

## Material and method:

### Extraction and culture of *Fusarium moniliforme*

*F. moniliforme* a fungus is found in the infected roots of rice plant. It causes bakanae and foot rot disease in rice plant resulting in enhanced growth and decreased yield.

For the culture of *F. moniliforme* infected portion of the root is cut and inoculate in petridish having potato sugarcane agar media. Placed this petri dish in BOD at temperature 250°C for 8 days. After 8days colonies came.

### Preparation of media

For media boiled 300g potato 1L distilled water for the extraction of starch and add 2ml sugarcane juice as C-source instead of glucose and 0.2g agar agar for solidification of media.

### PHA production in *Fusiform moniliforme*

PHA production was carried out in 500ml Erlenmeyer flask having 100ml liquid media contained different C:N ratios such as  $(\text{NH}_4)_2\text{SO}_4\text{FeSO}_4$  1g/l with different amount of sugarcane juice like 10ml, 20ml, 30ml or 20ml/l sugarcane juice with 2g, 4g, & 6g of  $(\text{NH}_4)_2\text{SO}_4\text{FeSO}_4$ . Incubations were carried out for 72 hours at 250rpm and 30°C temperature.

## Biomass & Polysaccharides estimation

After centrifugation of culture media cell biomass was collected and dried in airflow drier at temperature 70°C for biomass estimation. For polysaccharides estimation isopropanol was used. Polysaccharides were dried at 70°C upto a constant weight.

### PHA estimation:

For the extraction of PHA from biomass solvent extraction was used. The solvent employed for this was chloroform. As this method of extraction is very simple and effective to separate PHA granules from biomass. PHA obtained by this method was highly purified and without any degradation of PHA molecule.

### PHA production with varying concentration of carbon and nitrogen:

PHAs are mainly synthesized in the presence of excess of carbon with limiting amount of other nutrients like N, O, S, P etc. Production of PHA increases with increasing concentration of substrate that provides carbon and decreases with increasing concentration of substrate that provides nitrogen. Results are shown in the following table:

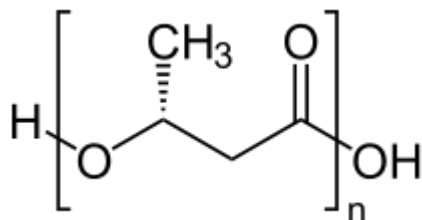
**Table1- Production of PHA with varying concentration of carbon and nitrogen**

Sugarcane juice (Carbon source) (ml/l)	Ammonium ferrous sulphate (Nitrogen source)(g/l)	Biomass (g dry wt/l)	PHA (g/l)
10	1	3.5	1
20	1	4	1.3
30	1	5	1.5
20	2	4	1
20	4	2.0	0.8
20	6	1	0.5

## Result and discussion

PHA synthesized by this method is mainly polyhydroxybutyrate(PHB).

### Structure of PHB



And its IR spectra and NMR spectra gives the following data:

**IR spectra** : 1098 (C-O), 1670 (C=O aliphatic), 2650 (CH<sub>2</sub>), 2927 (C-H stretching)

**NMR** : 1.45 (3H,d,-CH<sub>3</sub>), 1.874 (2H,d,-CH<sub>2</sub>-), 6.49(1H,m,-CH-)

The <sup>1</sup>H NMR were recorded in the indicated solvent on a Varian 500 MHz and 200 MHz spectrometer with TMS as internal standard. All chemical shifts (δ) were reported in ppm from internal TMS. Infrared spectra were recorded in KBr medium. The infrared spectra and proton

NMR data clearly suggest the molecule obtained is Polyhydroxybutyrate (PHB).

Use of sugarcane juice instead of sucrose/ glucose as carbon substrate is relatively cheap.

## Properties of PHA

- Water insoluble and relatively resistant to hydrolytic degradation. This differentiates PHB from most other currently available biodegradable, which are either water soluble or moisture sensitive.
- Good oxygen permeability.
- Good ultra-violet resistance but poor resistance to acids and bases.
- Soluble in chloroform and other chlorinated hydrocarbons.<sup>[8]</sup>
- Biocompatible and hence is suitable for medical applications.
- Melting point 175°C., and glass transition temperature 2°C.
- Tensile strength 40 MPa, close to that of polypropylene.
- Sinks in water (while polypropylene floats), facilitating its anaerobic biodegradation in sediments.
- Nontoxic.
- Less 'sticky' when melted, making it a potentially good material for clothing in the future.

## Biodegradability of PHA synthesized

PHB synthesized degrades in microbially active environment. Degradation occurs most rapidly in anaerobic sewage and slowest in seawater. Microorganisms colonize on the polymer surface and secrete enzyme that convert PHB into hydroxybutyrate (HB). These monomer units are then utilized by cell as carbon source.

Degradability rate depends on various factors like surface area, pH, temperature, pressure of other nutrients and moisture etc. In aerobic environment carbon dioxide and water are the end products of PHB degradation.

## Applications of PHB

The perspective area of PHB application is development of implanted medical devices for dental, craniomaxillofacial, orthopaedic, hernioplastic and skin surgery.

A number of potential medical devices on the base of PHB: (A) bioresorbable surgical suture; (B) biodegradable screws and plate for cartilage and bone fixation; (C) biodegradable membranes for periodontal treatment; (D) surgical meshes with PHB coating for hernioplastic surgery, wound coverings have been developed.

## Future outlook

PHAs production has drawn major attention due to its biocompatibility and biodegradability with conventional plastic. The production cost of PHAs through this method is less as compared to method in which glucose/sucrose used as carbon substrate. To produce it commercially researches are continuously going on by developing some mutant strains and using some alternative carbon sources.

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## References

[1] Witholt B, Kessler B. Perspectives of medium chain length poly (hydroxyalkanoates), a versatile set of bacterial bioplastics. *Current Opinion in Biotechnology*, **1999**, 279-285.

[2] Kunioka M, Kawaguchi Y, Doi Y. Production of biodegradable copolyesters of 3-hydroxybutyrate and 4-hydroxybutyrate by *Alcaligenes eutrophus*. *Applied Microbiology and Biotechnology*, **1989**, 569-573.

[3] Saito Y, Nakamura S, Hiramitsu M, Doi Y. Microbial synthesis and properties of poly(3-hydroxybutyrate-co-4-hydroxybutyrate). *Polymer International*, 1996, 169-174.

[4] Kellerhals MB, Kessler B, Witholt B, Tchouboukov A, Brandl H. Renewable long-chain fatty acids for production of biodegradable medium-chain-length polyhydroxyalkanoates (mcl-PHAs) at laboratory and pilot plant scales. *Macromolecules*, **2000**, 4690-4698.

[5] Lee SY, Wong HH, Choi JI, Lee SH, Lee SC, Han CS. Production of medium-chain-length polyhydroxyalkanoates by high cell-density cultivation *Pseudomonas putida* under phosphorus limitation. *Biotechnology and Bioengineering*, **2000**, 466-470.

[6] Yu P, Chua H, Huang AL, Ho KP. Conversion of industrial food wastes by *Alcaligenes latus* into polyhydroxyalkanoates. *Applied Biochemistry and Biotechnology*, **1999**, 445-454.

[7] Omar S, Rayes A, Eqaab A, Voß I, Steinbhel A. Optimization of cell growth and poly (3-hydroxybutyrate) accumulation on date syrup by a *Bacillus megaterium* strain. *Biotechnology Letters*, **2001**, 1119-1123.

[8] Chee JY, Tan Y, Samian MR, Sudesh K. Isolation and characterization of a Burkholderia sp. USM (JCM15050) capable of producing polyhydroxyalkanoate (PHA) from triglycerides, fatty acids and glycerols. *Journal of Polymer and the Environment*, **2010**, in press, doi: 10.1007/s10924-010-0204-1.

[9] Thakor N, Trivedi U, Patel KC. Biosynthesis of medium chain length poly (3-hydroxyalkanoates) (mcl-PHAs) by *Comamonas testosteroni* during cultivation on vegetable oils. *Bioresource Technology*, **2005**, 1843-1850.

[10] Yu J, Stahl H. Microbial utilization and biopolyester synthesis of bagasse hydrolysates. *Bioresource Technology*, **2008**, 8042-8048.

[11] Lee WH, Loo CY, Nomura CT, Sudesh K. Biosynthesis of polyhydroxyalkanoate copolymers from mixtures of plant oils and 3-hydroxyvalerate precursors. *Bioresource Technology*, **2008**, 6844-6851.

[12] Cavalheiro JMBT, de Almeida MCMD, Grandfils C, da Fonseca MMR. Poly (3-hydroxybutyrate) production by *Cupriavidus necator* using waste glycerol. *Process Biochemistry*, **2009**, 509-515.

[13] Loo CY, Lee WH, Tsuge T, Doi Y, Sudesh K. Biosynthesis and characterization of poly(3-

hydroxybutyrate-co-3-hydroxyhexanoate) from palm oil products in a *Wautersia eutropha* mutant. *Biotechnology Letters*, **2005**, 1405-1410.

[14]Fonseca GG, Antonio RV. Polyhydroxyalkanoates production by recombinant *Escherichia coli* harboring the structural genes of the polyhydroxyalkanoate synthases of *Ralstonia eutropha* and *Pseudomonas aeruginosa* using low cost substrate. *Journal of Applied Sciences*, **2006**, 1745-1750.

[15] Marsudi S, Unno H, Hori K. Palm oil utilization for the simultaneous production of polyhydroxyalkanoates and rhamnolipids by *Pseudomonas aeruginosa*. *Applied Microbiology and Biotechnology*, **2008**, 955-961.

[16] Fernadez D, Rodrauez E, Bassas M, Viñas M, Solonas AM, Llorens J, Marques AM, Manresa A. Agro-industrial oily wastes as substrates for PHA production by the new strain *Pseudomonas aeruginosa* NCIB 40045: Effect of culture conditions. *Biochemical Engineering*, **2005**, 159-167.

[17]Simon-Colin C, Raguenes G, Crassous P, Moppert X, Guezennec J. A novel mcl-PHA produced on coprah oil by *Pseudomonas guezennei* biovar. *tikehau*, isolated from a "kopara" mat of French Polynesia. *International Journal of Biological Macromolecules*, **2008**, 176-181.

[18] Pantazaki AA, Papaneophytou CP, Pritsa AG, Liakopoulou-Kyriakides M, Kyriakidis DA. Production of polyhydroxyalkanoates from whey by *Thermus thermophilus* HB8. *Process Biochemistry*, **2009**, 847-853.

[19]Kshama Lakshman & Tumkur Ramachandriah Shamala : Department of food and microbiology, Central food technological research institute , Mysore 570013, India, Enhanced biosynthesis of PHA in a mutant strain of *Rhizobium meliloti*, **2002**.

[20]. Shake flask studies and Fed batch fermented studies in *Alcaligenes latus* & *Alcaligenes eutrophus*: by middle east technical university, Department of biological sciences, Biotechnology research unit 06531 Ankara-Turkey

[21]. Ojumu, T.V., Yu, J. and Soloman, B.O.: Biodegradation of polymer engineering material department institute, P.M.B 611, Akure, Nigeria Hawaii Natural energy institute, university of Hawaii, Honolulu HI 96822, USA Department of chemical engineering, Obafemi Awolowo university, Nigeria, **2003**.

[22]. Robert G Kranz, Karen K. Gabbert, Terry A. Locke and Michael T. Madigan (1997): PHA production in *Rhodobacter capsulatus*. Department of biology, Washington University, St. Louis Missouri 63130& Department of microbiology, Southern Illinois university Carbondale Illinois 62901-6508

[23] Noda I. Process for recovering polyhydroxyalkanoates using air classification. United States Patent, **1998**, 849,854.

[24] Shinoka T, Shum-Tim D, Ma PX, Tanel RE, Isogai N, Langer R, Vacanti JP, Mayer JE. Creation of viable pulmonary artery autografts through tissue engineering. *The Journal of Thoracic Cardiovascular Surgery*. **1998**, 536-546.

[25] Keen I, Broota P, Rintoul L, Fredericks P, Trau M, Grøndahl L. Introducing amine functionalities on a poly (3-hydroxybutyrate-co-3-hydroxyvalerate) surface: comparing the use of ammonia plasma treatment and ethylenediamine aminolysis, Nanotechnology and Biomaterials Centre and School of Molecular and Microbial Sciences, The University of Queensland, Brisbane, Queensland, Australia , **2003**.